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Review Article

Role and Progress of DHA Dietary Supply in Brain Development of Maternal and Golden Periods

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Received: 02 July 2016 Accepted: 07 Aug 2016	Docosahexaenoic acid (DHA) dietary supplementation has highly increased in recent years. Sufficient DHA supplementation during gestation and early childhood is supremely clinical important. Some substantiation has been constructed for neurocognitive and brain benefits of supplementation with long chain polyunsaturated fatty acids (LC PUFA) such as DHA by the time before, during and after gestation. Acceptable in quality and quantity of DHA availability in the fetus and infant optimizes brain and retinal maturation development by affecting neurotransmitter pathways. DHA role as anti-inflammatory agent is significantly interceded over modulating of signaling pathways by directly binding to receptors or alteration in lipid raft formation and receptor presentation. Here, we intend to review the current progress on DHA supplementation, specifically brain development of fetus during pregnancy and infant, as a pharmacologic agent with both preventative and therapeutic endeavors. These findings provide benefits of routine DHA supplementation in pre-, during and post-gestation to improve neurodevelopmental outcomes in fetus, infant and early childhood. However, optimal composition of the supplement and dosing and treatment strategies still need to be determined toward advance nutrient for routine DHA dietary supply.
	Key words : DHA, LC PUFA, brain development, supplementation

1. INTRODUCTION

Docosahexaenoic acid (DHA) is a structural component of membrane cells particularly in the central nervous system (CNS)¹. DHA accumulation in the fetal brain occurs mainly during the last trimester of gestation and it will increase up to the end of the second year of life. Since the endogenous formation of DHA is relatively low, DHA intake may contribute to optimal brain development. The associations between DHA levels and brain development during the lifespan justify the indication of DHA role in differentiation to brain function for neuronal

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cell growth. Previous studies indicate that DHA may affect brain in early development in fetus, infancy and early childhood²⁻⁴.

DHA is the most unsaturated fatty acid, it is known as a building block of mammalian tissue⁵. DHA is normally appointed 22:6 3 or 22:6 3, indicates that it has 22 carbon atoms and six double bonds⁶. More than 95% of PUFA in mammalian tissues have double bonds in a specific arrangement known as 'homoallylic'⁷, where double bonds and methylene units (-CH₂-) alocate along the carbon chain⁸. Number of double bonds and the 3 (omega-3) position in PUFA completely specify the position of every double bond in the molecule. Thus, DHA shows consistency with most other mammalian unsaturated fatty acids and PUFA due to its structure feature its double bonds in the cis configuration^{9,10}.

Moreover, DHA is found in relatively high concentrations in all neural tissue, specifically in the brain¹ and retina¹¹. Early evidence showed that brain DHA concentration is highly constant across terrestrial species irrespective of the diversity of their natural diets, ranging over less than 50% ^{12,13}. In contrast, the liver in these same species have DHA concentrations that vary over 30-fold¹³. These observations are consistent with DHA supply as a limiting factor in adult brain size. More specifically, DHA's role cannot be completely replaced by the closest 6 PUFA molecular homologue, docosapentaenoic acid (DPA 6, 22:5 6), which is of identical structure to DHA with the exception of one missing double bond ^{14,15}.

Although DHA can be synthesized from its 3 precursor (ALA)¹⁶, studies have shown that DHA from the maternal diet is more efficient in source of neural tissue DHA than in an equivalent amount precursor of ALA ^{17, 18}. Precursor ALA only 10% could be converted into DHA^{19, 20}. DHA is one of the most variable fatty acids in human milk, its concentration being directly influenced by the amount of DHA in the maternal diet and parity²¹ in which the reported level of DHA in breast milk (by weight) of total fatty acids within the range of 0.06–1.4% ²². Current formula recommendations are between 0.2–0.5% DHA (g DHA/100 g of total fatty acids)^{23,24}.

Increasing attention over the last 20 years upon DHA in pregnancy and lactation for its role in brain development, as it accounts for over 10% of brain fatty acids is essential for infant brain development²⁵⁻²⁸. Infant brain doubles in weight, and the large brain to body weight ratio for infants (0.1) compared with adults (0.02) may put the infant at higher risk of deficiency in brain nutrients and energy^{29,30}. To anticipate this condition, mother diets in DHA intake is recommended in corresponding to the formation of neural synapses which are rich in DHA, neurite outgrowth, dendritic complexity and neurotransmitter metabolism are also highly reliant on DHA^{17,31,32}. This review will give wide aspects on role and progress of DHA dietary supply in brain development, especially in pre-conception, maternal and early childhood which is known as golden age period.

2. DHA BIOSYNTHESIS

Approximately sixty percent of the wet weight of the mammalian brain comprises lipids. Approximately 70% of the fatty acid amount is synthesize biochemically in mammalian cells, and 30% must be obtained from diet. Seafood, fish oils, and fortified foods are rich sources of 3 polyunsatuated fatty acids (omega-3 (3) PUFAs: eicosapentaenoic acid (EPA), docosapentaenoic acid (DPA 3), and docosahexaenoic acid (DHA), as well as cholesterol³³. DHA constitutes 14% of total fatty acids in the body and it is greatly concentrated in neuronal membranes and synapses³⁴⁻³⁶. Obviously, there are some evidence that DHA is required for neural and retinal cells that neither DPA 3 nor DPA 6 can replace it. In infancy period, DHA concentrations are much higher than DPA 3 in mother's breast milk and all tissues³⁷⁻⁴¹. It might probably not entirely concurrent due to breast milk has a similar chemical structure of long chain PUFA as it is found in neural tissue, which is the most rapidly growing tissue during infancy period⁴².

According to the general pathway of DHA biosynthesis, the major plant-based 3 is a-linolenic acid (ALA). ALA is considered as a precursor of 3 PUFA due to it is the only 3 found in seed oils and plants in the modern food supply. The primary fate of dietary ALA is oxidation for energy; however, a variable fraction of ALA is converted by a series of desaturations and elongations to the long chain 3 PUFA, finally resulting in DHA. Any of the intermediate 3 PUFA can be converted to DHA as well^{43,44}, as shown in Fig. 1.



Fig 1: DHA Biosynthesis Pathways

Previous studies reported that, animal maintains costly metabolic and genetic mechanisms to biosynthesize DHA when it is not in the diet. Recent data indicate that neurons can perform synthesis of DHA^{1,45,46}. However, DHA synthesis pathway appears to be very inefficient and essentially only results DPA 3 in adult people¹⁹. Mammalian biosynthesis of DHA is metabolically expensive⁴⁷. Tissue DHA is ultimately acquired from the diet either as DHA itself or any of some 3 PUFA which can be metabolically converted into DHA⁴⁸.

Literally, supplementation of ALA during pregnancy does not increase DHA in infant blood lipids⁴⁹. Similarly, during lactation, ALA supplementation has no detectable effect on the amount of DHA present in human milk⁵⁰⁻⁵². By adding high amount of ALA in infant formula also do not significantly increase the blood levels of DHA in formulafed infants⁵³. Thus, for both maternal and infant period, DHA diet supply is completely essential.

3. DHA ROLE IN PRECONCEPTION, MATERNITY AND AFTER BIRTH

DHA is essential structural building block of the CNS ⁵⁴⁻⁵⁷. DHA is a greatly importance due to it is specifically concentrated in the structural membrane lipids of the brain and visual elements of the retina⁵⁴⁻⁵⁷. The human brain undergoes rapid growth during the first 2 years of life^{58,59}, which there is a 12-fold increase of DHA concentration in tissue. The amount of DHA in the brain is continually turned over, recycled, and replenished by uptake from blood plasma⁶⁰. DHA accumulation is ~1.45 mg/day in infant brain while DHA turnover rate is ~3.75 mg/day⁶¹. The dietary requirement to compensate accretion and turnover is estimated around ~5.2 mg/day (1.45+3.75 mg/day)^{62,63}. DHA intake is suggested from either DHA dietary supply or from the conversion of DHA from ALA (-linolenic acid)¹.

As we know, the conversion of ALA to DHA in the body is very limited $(<1\%)^{64-66}$. To maintain homeostasis of DHA (5.2 mg/day) in the brain, over 200 mg/day of dietary DHA is needed (Fig.2), assuming that only 1.7% of performed dietary DHA reaches the brain ⁶⁷⁻⁶⁹ and the other source is provided from conversion of ALA to DHA⁶¹. Furthermore, earlier infant will need higher the level of DHA than maternal term, thus contributing the optimal neurodevelopment could be optimum⁷⁰⁻⁷³.

Widely consensus statements recommend minimal diet is 200 mg per day of DHA for maternity and lactating women (Fig.2)^{3,4}. In fact, many lactating women are only receiving around 25% of this recommended amount^{13,39}. The breast milk DHA concentration was 4–6 times lower in the current baseline sample compared with those with the highest fish intake^{18,43} and 1.5 times lower than the worldwide mean¹⁸. Supplementation was able to achieve a concentration similar (low dose) or slightly higher (high dose) than the worldwide mean, suggesting that supplementation is necessary for those with low dietary intake. Thus, DHA supplementation results in an increase in breast milk and maternal DHA at that levels would reflect adequate dietary intake and beneficially impact fatty acid ratios in infants important for brain development.



Fig 2: Recommended dietary DHA amount in preconception, maternity and after birth

4. DHA DIETARY SOURCES

The type of DHA sources may also affect neurocognitive function in infants and childhood. The majority of studies have considered both neurocognitive function and growth and development have used infant formulas for maternal and infant term that contain DHA obtained from DHA from fish oil ^{72,74}, eggs^{75,76} and micro algae have been considered. This is noteworthy for both maternal⁷⁷⁻⁸⁰ and infant term^{81,82}. These DHA sources genuinely important due to it has positive impact in neural growth.

 Table 1: Source of Natural DHA from fishes, plants and marine organisms

Source of Fatty AcidTotal		DHA	Energy	Reference
(FA)	PUFA	(g/100	g(kcal)	
	(g/100	gtotal F	A)	
	total FA)			
Fish				÷
Trout	1.831	0.52	128	83
Salmon	3.256	1.429	155	83
Catfish	0.636	0.136	89	83
Tuna	*N/A	0.223	N/A	16
Sardine	N/A	0.509	N/A	16
Plant				
Canola (Rapeseed)	30	0	540	84
Flaxseed	56.31	0	N/A	85
Corn oil	0.94	0	N/A	44
Marine organism				
Microalgae				
(Crypthecodinium	51.8	27	N/A	86
cohnii)	N/A	7	N/A	87
Krill				

*N/A: Not Available

Fish consumption is considered as surrogate for omega-3 fatty acid consumption, which associates with cognitive outcomes in pregnant women ^{88,89}. Children from mothers with no seafood consumption were at greatest risk of adverse or suboptimal verbal IQ. Overall, consumption of more than 340 g seafood per week was beneficial for the child's neurodevelopment.

The most available source of DHA is fish, especially marine fatty fish, thus food enriched with fish oil is excellent source. DHA is naturally found in human milk and now included in almost all commercially available infant formulas. In fetus, DHA and other LC PUFA is derived from the mother by placental transfer and after birth by the infant diet (human milk or formula). There is a specific transport mechanism for the placental transfer of DHA from the mother to the fetus⁹⁰. The levels of DHA in human milk is increased by increasing maternal intake⁵². Human milk concentration of DHA vary widely and range from 0.17 to 0.99%, with the highest levels in Japanese milk and the lowest levels in Canadian and US samples ⁹¹. However, only modest variations in ARA levels in human milk are observed (0.36-0.49%) across different populations⁹¹.

Recent studies have shown that DHA status is influenced not only by diet, but also by genetic variants, single nucleotide polymorphisms (SNPs) in the fatty acid desaturates (FADS)⁹²⁻⁹⁵. The genetically explained variability of red blood cell DHA and ARA levels were very low (0.51% and 1.13%, respectively)⁹⁴. Furthermore, the effect of FADS

genetic variants on LC PUFA metabolism, specifically ARA, appears to vary at the population level⁹⁶.

Moreover, there has been a trend in increased consumption of refined vegetable oils, such as soybean and canola, as the sources of good fat in the diet and subsequent decreased animal fat consumption. However, it has resulted in an overall increase in 6 in the diet as well as decreased plant sources of 3, including walnuts and flax, suggesting a greater importance for intake of DHA diet 97,98 . These dietary changes, taken together with the overall shift in available dietary fatty acids over the past several decades have resulted in a significant increase value in the ratio of 6:3 fatty acids to 15–20:1, meanwhile ratios closer to 2–4:1 have been recommended for health $^{99-101}$.

ALA is practically the sole 3 PUFA in plant foods, and it is found primarily in components of leafy foods such as lettuce, spinach, cabbage, and Brussels sprouts, particularly in dry climate regions¹⁰². Animal metabolism is therefore required to synthesize most DHA from plant foods that cannot be obtained from animal foods with preformed DHA⁷. Though DHA can be a predominant fatty acid in specialized tissue. Animal part other than the brain specifically provide only few DHA accumulation. Thus DHA diet in food consumption will provide at least 35% DHA which can be used for energy ^{24, 103}.

5. CONCLUSION

In conclusion, the essential role of DHA in neurological development during the pre- and postnatal periods have been defined. Overall, evidence for the therapeutic advantages of LCPUFAs supplementation is strong. Of primary concern are the low levels of dietary DHA consumption in women of child bearing age from westernized countries and that sufficient quantities of DHA may not be available for optimal neurological development and/or immunological support to the developing infant. This is especially important in the preterm or small for gestational age infant. Clinical recommendations exist for pregnant and lactating women which should be focused on optimizing supplementation strategies to provide optimal outcomes for all childhood.

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9. REFERENCES

- 1. Bradbury J. Docosahexaenoic acid (DHA): an ancient nutrient for the modern human brain. Nutrients 2011;3(5):529-54.
- Lauritzen L, Brambilla P, Mazzocchi A, Harslof LB, Ciappolino V, Agostoni C. DHA Effects in Brain Development and Function. Nutrients 2016;8(1).
- 3. Prado EL, Dewey KG. Nutrition and brain development in early life. Nutr Rev 2014;72(4):267-84.

- 4. Innis SM. Dietary (n-3) fatty acids and brain development. J Nutr 2007;137(4):855-9.
- Bourre JM. [Biochemistry of brain lipids (especially fatty acids). In situ synthesis and exogenous origin during development. Various aspects of nutritional effects]. Reprod Nutr Dev 1982;22(1B):179-91.
- Tassoni D, Kaur G, Weisinger RS, Sinclair AJ. The role of eicosanoids in the brain. Asia Pac J Clin Nutr 2008;17 Suppl 1:220-8.
- Brenna JT, Carlson SE. Docosahexaenoic acid and human brain development: evidence that a dietary supply is needed for optimal development. J Hum Evol 2014; 77:99-106.
- 8. Slawson V, Stein RA. Comparative autoxidative susceptibility of fatty esters with O-6 methylene-interrupted double bonds. Lipids 1970;5(8):713-7.
- Muskiet FAJ. Pathophysiology and Evolutionary Aspects of Dietary Fats and Long-Chain Polyunsaturated Fatty Acids across the Life Cycle. In: Montmayeur JP, le Coutre J, editors. Fat Detection: Taste, Texture, and Post Ingestive Effects, Frontiers in Neuroscience. Boca Raton (FL)2010.
- Yazdi S, Stein M, Elinder F, Andersson M, Lindahl E. The Molecular Basis of Polyunsaturated Fatty Acid Interactions with the Shaker Voltage-Gated Potassium Channel. PLoS Comput Biol 2016;12(1):e1004704.
- Zemski Berry KA, Gordon WC, Murphy RC, Bazan NG. Spatial organization of lipids in the human retina and optic nerve by MALDI imaging mass spectrometry. J Lipid Res 2014;55(3):504-15.
- Mantzioris E, James MJ, Gibson RA, Cleland LG. Differences exist in the relationships between dietary linoleic and alpha-linolenic acids and their respective long-chain metabolites. Am J Clin Nutr 1995;61(2):320-4.
- 13. Crawford MA, Casperd NM, Sinclair AJ. The long chain metabolites of linoleic avid linolenic acids in liver and brain in herbivores and carnivores. Comp Biochem Physiol B 1976;54(3):395-401.
- unnane SC. Docosahexaenoic acid and human brain evolution: missing the forest for the trees--comments by Cunnane. Br J Nutr 2007;97(5):1021-2; discussion 1025.
- 15. Byelashov OA, Sinclair AJ, Kaur G. Dietary sources, current intakes, and nutritional role of omega-3 docosapentaenoic acid. Lipid Technol 2015;27(4):79-82.
- 16. Abedi E, Sahari MA. Long-chain polyunsaturated fatty acid sources and evaluation of their nutritional and functional properties. Food Sci Nutr 2014;2(5):443-63.
- 17. Sherry CL, Oliver JS, Marriage BJ. Docosahexaenoic acid supplementation in lactating women increases breast milk and plasma docosahexaenoic acid concentrations and alters infant omega 6:3 fatty acid

- Int J Pharma Res Health Sci. 2016; 4 (4): 1268–1275 ratio. Prostaglandins Leukot Essent Fatty Acids 2015;95:63-9.
- 18. Liu L, Bartke N, Van Daele H, Lawrence P, Qin X, Park HG, Kothapalli K, Windust A, Bindels J, Wang Z and others. Higher efficacy of dietary DHA provided as a phospholipid than as a triglyceride for brain DHA accretion in neonatal piglets. J Lipid Res 2014;55(3):531-9.
- Brenna JT, Salem N, Jr., Sinclair AJ, Cunnane SC. alpha-Linolenic acid supplementation and conversion to n-3 long-chain polyunsaturated fatty acids in humans. Prostaglandins Leukot Essent Fatty Acids 2009;80(2-3):85-91.
- 20. Goyens PL, Spilker ME, Zock PL, Katan MB, Mensink RP. Conversion of alpha-linolenic acid in humans is influenced by the absolute amounts of alpha-linolenic acid and linoleic acid in the diet and not by their ratio. Am J Clin Nutr 2006;84(1):44-53.
- Bopp M, Lovelady C, Hunter C, Kinsella T. Maternal diet and exercise: effects on long-chain polyunsaturated fatty acid concentrations in breast milk. J Am Diet Assoc 2005;105(7):1098-103.
- 22. Antonakou A, Skenderi KP, Chiou A, Anastasiou CA, Bakoula C, Matalas AL. Breast milk fat concentration and fatty acid pattern during the first six months in exclusively breastfeeding Greek women. Eur J Nutr 2013;52(3):963-73.
- 23. Kelishadi R, Hadi B, Iranpour R, Khosravi-Darani K, Mirmoghtadaee P, Farajian S, Poursafa P. A study on lipid content and fatty acid of breast milk and its association with mother's diet composition. J Res Med Sci 2012;17(9):824-7.
- 24. Kris-Etherton PM, Grieger JA, Etherton TD. Dietary reference intakes for DHA and EPA. Prostaglandins Leukot Essent Fatty Acids 2009;81(2-3):99-104.
- 25. Dubnov-Raz G, Finkelstein Y, Koren G. Omega-3 fatty acid supplementation during pregnancy: for mother, baby, or neither? Can Fam Physician 2007;53(5):817-8.
- 26. Furuhjelm C, Jenmalm MC, Falth-Magnusson K, Duchen K. Th1 and Th2 chemokines, vaccine-induced immunity, and allergic disease in infants after maternal omega-3 fatty acid supplementation during pregnancy and lactation. Pediatr Res 2011;69(3):259-64.
- 27. Greenberg JA, Bell SJ, Ausdal WV. Omega-3 Fatty Acid supplementation during pregnancy. Rev Obstet Gynecol 2008;1(4):162-9.
- Warstedt K, Furuhjelm C, Duchen K, Falth-Magnusson K, Fageras M. The effects of omega-3 fatty acid supplementation in pregnancy on maternal eicosanoid, cytokine, and chemokine secretion. Pediatr Res 2009;66(2):212-7.
- 29. Fontana R, Della Torre S. The Deep Correlation between Energy Metabolism and Reproduction: A View on the Effects of Nutrition for Women Fertility. Nutrients 2016;8(2):87.

- 30. Nutrition and reproduction in women. Hum Reprod Update 2006;12(3):193-207.
- 31. Wurtman RJ, Cansev M, Sakamoto T, Ulus I. Nutritional modifiers of aging brain function: use of uridine and other phosphatide precursors to increase formation of brain synapses. Nutr Rev 2010;68 Suppl 2:S88-101.
- 32. Bazan NG, Molina MF, Gordon WC. Docosahexaenoic acid signalolipidomics in nutrition: significance in aging, neuroinflammation, macular degeneration, Alzheimer's, and other neurodegenerative diseases. Annu Rev Nutr 2011;31:321-51.
- 33. Du J, Zhu M, Bao H, Li B, Dong Y, Xiao C, Zhang GY, Henter I, Rudorfer M, Vitiello B. The Role of Nutrients in Protecting Mitochondrial Function and Neurotransmitter Signaling: Implications for the Treatment of Depression, PTSD, and Suicidal Behaviors. Crit Rev Food Sci Nutr 2014:0.
- 34. Morse NL. Benefits of docosahexaenoic acid, folic acid, vitamin D and iodine on foetal and infant brain development and function following maternal supplementation during pregnancy and lactation. Nutrients 2012;4(7):799-840.
- 35. Joensen AM, Schmidt EB, Dethlefsen C, Johnsen SP, Tjonneland A, Rasmussen LH, Overvad K. Dietary intake of total marine n-3 polyunsaturated fatty acids, eicosapentaenoic acid, docosahexaenoic acid and docosapentaenoic acid and the risk of acute coronary syndrome - a cohort study. Br J Nutr 2010;103(4):602-7.
- 36. Green KN, Martinez-Coria H, Khashwji H, Hall EB, Yurko-Mauro KA, Ellis L, LaFerla FM. Dietary docosahexaenoic acid and docosapentaenoic acid ameliorate amyloid-beta and tau pathology via a mechanism involving presenilin 1 levels. J Neurosci 2007;27(16):4385-95.
- Saccone G, Saccone I, Berghella V. Omega-3 longchain polyunsaturated fatty acids and fish oil supplementation during pregnancy: which evidence? J Matern Fetal Neonatal Med 2016;29(15):2389-97.
- 38. Kemse NG, Kale AA, Joshi SR. A combined supplementation of omega-3 fatty acids and micronutrients (folic acid, vitamin B12) reduces oxidative stress markers in a rat model of pregnancy induced hypertension. PLoS One 2014;9(11):e111902.
- 39. Khaire A, Rathod R, Kemse N, Kale A, Joshi S. Supplementation with omega-3 fatty acids during gestation and lactation to a vitamin B12-deficient or supplemented diet improves pregnancy outcome and metabolic variables in Wistar rats. Reprod Fertil Dev 2015;27(2):341-50.
- Larque E, Gil-Sanchez A, Prieto-Sanchez MT, Koletzko B. Omega 3 fatty acids, gestation and pregnancy outcomes. Br J Nutr 2012;107 Suppl 2:S77-84.

- 41. Coletta JM, Bell SJ, Roman AS. Omega-3 Fatty acids and pregnancy. Rev Obstet Gynecol 2010;3(4):163-71.
- Arterburn LM, Hall EB, Oken H. Distribution, interconversion, and dose response of n-3 fatty acids in humans. Am J Clin Nutr 2006;83(6 Suppl):1467S-1476S.
- Rajaram S. Health benefits of plant-derived alphalinolenic acid. Am J Clin Nutr 2014;100 Suppl 1:443S-8S.
- 44. Hirschi KD. Nutrient biofortification of food crops. Annu Rev Nutr 2009;29:401-21.
- 45. Weiser MJ, Butt CM, Mohajeri MH. Docosahexaenoic Acid and Cognition throughout the Lifespan. Nutrients 2016;8(2):99.
- 46. Dyall SC. Long-chain omega-3 fatty acids and the brain: a review of the independent and shared effects of EPA, DPA and DHA. Front Aging Neurosci 2015;7:52.
- Ruiz-Lopez N, Usher S, Sayanova OV, Napier JA, Haslam RP. Modifying the lipid content and composition of plant seeds: engineering the production of LC-PUFA. Appl Microbiol Biotechnol 2015;99(1):143-54.
- 48. Kitessa SM, Abeywardena M, Wijesundera C, Nichols PD. DHA-containing oilseed: a timely solution for the sustainability issues surrounding fish oil sources of the health-benefitting long-chain omega-3 oils. Nutrients 2014;6(5):2035-58.
- 49. de Groot RH, Hornstra G, van Houwelingen AC, Roumen F. Effect of alpha-linolenic acid supplementation during pregnancy on maternal and neonatal polyunsaturated fatty acid status and pregnancy outcome. Am J Clin Nutr 2004;79(2):251-60.
- 50. Liu G, Ding Z, Li X, Chen X, Wu Y, Xie L. Relationship between polyunsaturated fatty acid levels in maternal diets and human milk in the first month post-partum. J Hum Nutr Diet 2015.
- 51. Iranpour R, Kelishadi R, Babaie S, Khosravi-Darani K, Farajian S. Comparison of long chain polyunsaturated fatty acid content in human milk in preterm and term deliveries and its correlation with mothers' diet. J Res Med Sci 2013;18(1):1-5.
- 52. Innis SM. Polyunsaturated fatty acids in human milk: an essential role in infant development. Adv Exp Med Biol 2004;554:27-43.
- 53. Ponder DL, Innis SM, Benson JD, Siegman JS. Docosahexaenoic acid status of term infants fed breast milk or infant formula containing soy oil or corn oil. Pediatr Res 1992;32(6):683-8.
- 54. Innis SM. Dietary omega 3 fatty acids and the developing brain. Brain Res 2008;1237:35-43.
- 55. Yamashita A, Kawana K, Tomio K, Taguchi A, Isobe Y, Iwamoto R, Masuda K, Furuya H, Nagamatsu T, Nagasaka K and others. Increased tissue levels of omega-3 polyunsaturated fatty acids prevents pathological preterm birth. Sci Rep 2013;3:3113.

- 56. O'Brien JS, Sampson EL. Fatty acid and fatty aldehyde composition of the major brain lipids in normal human gray matter, white matter, and myelin. J Lipid Res 1965;6(4):545-51.
- 57. Seo T, Blaner WS, Deckelbaum RJ. Omega-3 fatty acids: molecular approaches to optimal biological outcomes. Curr Opin Lipidol 2005;16(1):11-8.
- Chase HP, Canosa CA, Dabiere CS, Welch NN, O'Brien D. Postnatal undernutrition and human brain development. J Ment Defic Res 1974;18(4):355-66.
- Dobbing J. Undernutrition and the developing brain. The relevance of animal models to the human problem. Bibl Nutr Dieta 1972(17):35-46.
- 60. Innis SM. Essential fatty acid transfer and fetal development. Placenta 2005;26 Suppl A:S70-5.
- Hadley KB, Ryan AS, Nelson EB, Salem N. An assessment of dietary docosahexaenoic acid requirements for brain accretion and turnover during early childhood. World Rev Nutr Diet 2009;99:97-104.
- 62. Sievers E. Nutrient requirements for preterm infant formulas-molybdenum. J Nutr 2003;133(1):236-7.
- 63. Klein CJ. Nutrient requirements for preterm infant formulas. J Nutr 2002;132(6 Suppl 1):1395S-577S.
- 64. Pawlosky RJ, Hibbeln JR, Lin Y, Goodson S, Riggs P, Sebring N, Brown GL, Salem N, Jr. Effects of beef- and fish-based diets on the kinetics of n-3 fatty acid metabolism in human subjects. Am J Clin Nutr 2003;77(3):565-72.
- Goyens PL, Spilker ME, Zock PL, Katan MB, Mensink RP. Compartmental modeling to quantify alphalinolenic acid conversion after longer term intake of multiple tracer boluses. J Lipid Res 2005;46(7):1474-83.
- 66. Burdge G. Alpha-linolenic acid metabolism in men and women: nutritional and biological implications. Curr Opin Clin Nutr Metab Care 2004;7(2):137-44.
- 67. Bowen RA, Clandinin MT. High dietary 18:3n-3 increases the 18:3n-3 but not the 22:6n-3 content in the whole body, brain, skin, epididymal fat pads, and muscles of suckling rat pups. Lipids 2000;35(4):389-94.
- Su HM, Bernardo L, Mirmiran M, Ma XH, Nathanielsz PW, Brenna JT. Dietary 18:3n-3 and 22:6n-3 as sources of 22:6n-3 accretion in neonatal baboon brain and associated organs. Lipids 1999;34 Suppl:S347-50.
- 69. Bowen RA, Clandinin MT. Maternal dietary 22 : 6n-3 is more effective than 18 : 3n-3 in increasing the 22 : 6n-3 content in phospholipids of glial cells from neonatal rat brain. Br J Nutr 2005;93(5):601-11.
- 70. Makrides M. DHA supplementation during the perinatal period and neurodevelopment: Do some babies benefit more than others? Prostaglandins Leukot Essent Fatty Acids 2013;88(1):87-90.
- 71. Davis-Bruno K, Tassinari MS. Essential fatty acid supplementation of DHA and ARA and effects on neurodevelopment across animal species: a review of

- Int J Pharma Res Health Sci. 2016; 4 (4): 1268–1275 the literature. Birth Defects Res B Dev Reprod Toxicol 2011;92(3):240-50.
- 72. Makrides M, Gibson RA, McPhee AJ, Yelland L, Quinlivan J, Ryan P. Effect of DHA supplementation during pregnancy on maternal depression and neurodevelopment of young children: a randomized controlled trial. JAMA 2010;304(15):1675-83.
- 73. Uauy R, Hoffman DR, Mena P, Llanos A, Birch EE. Term infant studies of DHA and ARA supplementation on neurodevelopment: results of randomized controlled trials. J Pediatr 2003;143(4 Suppl):S17-25.
- 74. Molloy CS, Stokes S, Makrides M, Collins CT, Anderson PJ, Doyle LW. Long-term effect of high-dose supplementation with DHA on visual function at school age in children born at <33 wk gestational age: results from a follow-up of a randomized controlled trial. Am J Clin Nutr 2016;103(1):268-75.
- 75. Fewtrell MS, Morley R, Abbott RA, Singhal A, Isaacs EB, Stephenson T, MacFadyen U, Lucas A. Doubleblind, randomized trial of long-chain polyunsaturated fatty acid supplementation in formula fed to preterm infants. Pediatrics 2002;110(1 Pt 1):73-82.
- 76. Lucas A, Stafford M, Morley R, Abbott R, Stephenson T, MacFadyen U, Elias-Jones A, Clements H. Efficacy and safety of long-chain polyunsaturated fatty acid supplementation of infant-formula milk: a randomised trial. Lancet 1999;354(9194):1948-54.
- 77. Clandinin MT, Van Aerde JE, Merkel KL, Harris CL, Springer MA, Hansen JW, Diersen-Schade DA. Growth and development of preterm infants fed infant formulas containing docosahexaenoic acid and arachidonic acid. J Pediatr 2005;146(4):461-8.
- 78. Carvajal JA. Docosahexaenoic acid supplementation early in pregnancy may prevent deep placentation disorders. Biomed Res Int 2014;2014:526895.
- 79. Westerberg AC, Schei R, Henriksen C, Smith L, Veierod MB, Drevon CA, Iversen PO. Attention among very low birth weight infants following early supplementation with docosahexaenoic and arachidonic acid. Acta Paediatr 2011;100(1):47-52.
- 80. Henriksen C, Haugholt K, Lindgren M, Aurvag AK, Ronnestad A, Gronn M, Solberg R, Moen A, Nakstad B, Berge RK and others. Improved cognitive development among preterm infants attributable to early supplementation of human milk with docosahexaenoic acid and arachidonic acid. Pediatrics 2008;121(6):1137-45.
- Birch EE, Garfield S, Hoffman DR, Uauy R, Birch DG. A randomized controlled trial of early dietary supply of long-chain polyunsaturated fatty acids and mental development in term infants. Dev Med Child Neurol 2000;42(3):174-81.
- 82. Drover JR, Hoffman DR, Castaneda YS, Morale SE, Garfield S, Wheaton DH, Birch EE. Cognitive function in 18-month-old term infants of the DIAMOND study: a

randomized, controlled clinical trial with multiple dietary levels of docosahexaenoic acid. Early Hum Dev 2011;87(3):223-30.

- Gebauer SK, Psota TL, Harris WS, Kris-Etherton PM. n-3 fatty acid dietary recommendations and food sources to achieve essentiality and cardiovascular benefits. Am J Clin Nutr 2006;83(6 Suppl):1526S-1535S.
- 84. Jones PJ, Senanayake VK, Pu S, Jenkins DJ, Connelly PW, Lamarche B, Couture P, Charest A, Baril-Gravel L, West SG and others. DHA-enriched high-oleic acid canola oil improves lipid profile and lowers predicted cardiovascular disease risk in the canola oil multicenter randomized controlled trial. Am J Clin Nutr 2014;100(1):88-97.
- Sahar Soltan, Abdel Magied Soltan, 2012, The Effects of Varieties Sources of Omega-3 Fatty Acids on Diabetes in Rats. Food and Nutrition Sciences, 3: 1404-1412
- Sprague M, Dick JR, Tocher DR. Impact of sustainable feeds on omega-3 long-chain fatty acid levels in farmed Atlantic salmon, 2006-2015. Sci Rep 2016;6:21892.
- 87. Burri L, Johnsen L. Krill products: an overview of animal studies. Nutrients 2015;7(5):3300-21.
- 88. Julvez J, Mendez M, Fernandez-Barres S, Romaguera D, Vioque J, Llop S, Ibarluzea J, Guxens M, Avella-Garcia C, Tardon A and others. Maternal Consumption of Seafood in Pregnancy and Child Neuropsychological Development: A Longitudinal Study Based on a Population With High Consumption Levels. Am J Epidemiol 2016;183(3):169-82.
- Hibbeln JR, Davis JM, Steer C, Emmett P, Rogers I, Williams C, Golding J. Maternal seafood consumption in pregnancy and neurodevelopmental outcomes in childhood (ALSPAC study): an observational cohort study. Lancet 2007;369(9561):578-85.
- Koletzko B, Larque E, Demmelmair H. Placental transfer of long-chain polyunsaturated fatty acids (LC-PUFA). J Perinat Med 2007;35 Suppl 1:S5-11.
- Yuhas R, Pramuk K, Lien EL. Human milk fatty acid composition from nine countries varies most in DHA. Lipids 2006;41(9):851-8.
- 92. Koletzko B, Demmelmair H, Schaeffer L, Illig T, Heinrich J. Genetically determined variation in polyunsaturated fatty acid metabolism may result in different dietary requirements. Nestle Nutr Workshop Ser Pediatr Program 2008;62:35-44; discussion 44-9.
- 93. Lattka E, Koletzko B, Zeilinger S, Hibbeln JR, Klopp N, Ring SM, Steer CD. Umbilical cord PUFA are determined by maternal and child fatty acid desaturase (FADS) genetic variants in the Avon Longitudinal Study of Parents and Children (ALSPAC). Br J Nutr 2013;109(7):1196-210.
- 94. Koletzko B, Lattka E, Zeilinger S, Illig T, Steer C. Genetic variants of the fatty acid desaturase gene cluster

- predict amounts of red blood cell docosahexaenoic and other polyunsaturated fatty acids in pregnant women: findings from the Avon Longitudinal Study of Parents and Children. Am J Clin Nutr 2011;93(1):211-9.
- 95. zehak P, Heinrich J, Klopp N, Schaeffer L, Hoff S, Wolfram G, Illig T, Linseisen J. Evidence for an association between genetic variants of the fatty acid desaturase 1 fatty acid desaturase 2 (FADS1 FADS2) gene cluster and the fatty acid composition of erythrocyte membranes. Br J Nutr 2009;101(1):20-6.
- 96. Mathias RA, Sergeant S, Ruczinski I, Torgerson DG, Hugenschmidt CE, Kubala M, Vaidya D, Suktitipat B, Ziegler JT, Ivester P and others. The impact of FADS genetic variants on omega6 polyunsaturated fatty acid metabolism in African Americans. BMC Genet 2011;12:50.
- 97. Kris-Etherton PM, Taylor DS, Yu-Poth S, Huth P, Moriarty K, Fishell V, Hargrove RL, Zhao G, Etherton TD. Polyunsaturated fatty acids in the food chain in the United States. Am J Clin Nutr 2000;71(1 Suppl):179S-88S.
- Kris-Etherton PM, Fleming JA. Emerging nutrition science on fatty acids and cardiovascular disease: nutritionists' perspectives. Adv Nutr 2015;6(3):326S-37S.
- 99. Powell ML, Pegues MA, Szalai AJ, Ghanta VK, D'Abramo LR, Watts SA. Effects of the Dietary omega3:omega6 Fatty Acid Ratio on Body Fat and Inflammation in Zebrafish (Danio rerio). Comp Med 2015;65(4):289-94.
- 100.Simopoulos AP. Importance of the ratio of omega-6/omega-3 essential fatty acids: evolutionary aspects. World Rev Nutr Diet 2003;92:1-22.
- 101.Simopoulos AP. The importance of the ratio of omega-6/omega-3 essential fatty acids. Biomed Pharmacother 2002;56(8):365-79.
- 102. Pereira C, Li D, Sinclair AJ. The alpha-linolenic acid content of green vegetables commonly available in Australia. Int J Vitam Nutr Res 2001;71(4):223-8.
- 103.Rahmawaty S, Charlton K, Lyons-Wall P, Meyer BJ. Dietary intake and food sources of EPA, DPA and DHA in Australian children. Lipids 2013; 48(9):869-77.

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