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Original Article

The Performance of Nanostructured Lipid Carrier (NLC) Incorporated Transdermal Patch Coenzym Q10 : Effect of Lipid Ratio as Drug Reservoir and HPMC 606 as Rate Controlling Membrane

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ABSTRACT

Received:19 Sep 2018 Accepted:20 Oct 2018 Coenzyme Q10 is a strong antioxidant. Coenzyme Q10 can improve cellular dynamics in human body and play an effective role in preventing skin aging, keratinization and DNA oxidative. Coenzyme Q10 NLC system with different lipid ratio 70:30 (cetyl palmitate and alpha tocopheryl acetate) used as drug reservoir dan hydroxypropyl methylcellulose 606 (HPMC 606) as rate controlling membrane. NLC coenzyme Q10 as drug reservoir were prepared using high shear homogenization method. Transdermal patch was using membrane type with HPMC 606 as rate controlling membrane. This research was investigate the effect of concentration of HPMC 10%, 15% and 20% on the characteristics coenzyme Q10 transdermal patch. The prepared transdermal patches were evaluated for thickness, weight variation, moisture content, drug content and drug homogenity which were found to $1.98 \pm$ 0.003 mm, 1.422 ± 0.003 g, 8.441 ± 0.077 %, 96.90 ± 0.92 and 97.75 ± 1.78 %, respectively. All independent variables had no significant effect on the dependent variables (p-values >0.05) using one way ANOVA, except the weight and moisture content patches.

Key words: Coenzyme Q10, Hydroxypropyl methyl cellulose606, Nanostructured lipid carrier, Characteristic of transdermal patch

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1. INTRODUCTION

Coenzyme Q10 has functions of providing and preventing the skin. Reactive oxygen species can be restrained and the peroxidation form of lipid would be slowed and protected by coenzyme Q10. However, coenzyme Q10 has limited due to

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solubility in water is very low (4 ng/ml), causing low bioavailability and permeability of the oral administration¹. NLCs were developed as improvements over Solid Lipid Nanoparticles (SLNs). SLNs are composed of a solid lipid, whereas the NLC lipid phase contains both solid and liquid lipids. NLC increases the amount of liquid lipid such that the nanoparticles form a non-standard shape and stack, resulting in a non-perfect lattice and forming an amorphous structure. As increasing the liquid lipid can enhance the solubility of active substances, and active substances can help with better encapsulation NLCs lipid material shows more application potential. NLCs augment drug stability and loading capacity, and reduce drug leakage during storage². NLC can penetrate the skin and pass through the barrier of the stratum corneum by intercellular mechanisms, they move according to the osmotic gradient to a deep hydrated layer. Solid lipid in NLC system have been widely used as drug reservoir for the controlled release of active agents ^{3, 4}.

Transdermal patches offer an attractive route of administration for patients who are unable to swallow oral medications and to avoid the need for pain or risks associated with intravenous administration⁵. Patches may also allow for less frequent dosing and improved disadvantages 'drug expulsions' of NLC. Permeation enhancement is primarily due to small size and swelling of stratum corneum by an increase in skin hydration caused by the occlusive film of NLC². Enhancement of transdermal drug permeation of coenzyme Q10 using NLC have been reported, studied that coenzyme Q10 using solid lipid cetyl palmitate and alpha tocopherol acetate ratio 70:30 (% w/w) showed good characteristics, higher % EE, good penetration, controlled release, and stable during 90 days storage⁶.

The aim of this study was to formulate coenzyme Q10 NLC system with different lipid ratio 70:30 (cetyl palmitate and alpha tocopheryl acetate) used as drug reservoir and HPMC 606 as rate controlling membrane and evaluate the characteristics coenzyme Q10 patch to enhance skin penetration.

2. MATERIALS AND METHODS

Materials

Coenzyme Q10 was obtained from Xi'an Future Biotechnology, Co., Ltd., cetyl palmitate Cutina®CP (Cognis Chemical Care), alpha tocopherol acetate (Xinchang pharma), propylene glycol (Dow Chemical Pacific), Tween 80 (KAO.,Ltd), HPMC 606 (hydroxy propyl methyl cellulose606) (Wuhan Senwayer Century Chemical Co., Ltd), Cetostearyl alcohol and menthol (Bratachem.,Ltd), Ethanol pro analysis, NaOH (sodium hydroxide) and NaH2PO4 (natrium dihydrogen phosphate) pro analysis (Merck). Aqua demineralized (Bratachem.,Ltd), Backing layer was gifted by Pharmaceutic laboratory of Airlangga University. All other chemicals were of analytical grade.

Preparation Coenzyme Q10 NLC System

Coenzyme Q10- NLC was produced by hot High Pressure Homogenization (HPH). Briefly, after melting the lipid phase (cetyl palmitate : alpha tocopheryl acetate) ratio 70:30, at a temperature 65°C, coenzyme Q10-NLC was added until thoroughly dissolved and the mixture was immediately dispersed in a hot surfactant solution using an Ultra-Turrax High Shear Homogenizer IKA T-25 at 20.000 rpm for 8 minutes. The pre emulsion was further processed by high pressure homogenizer at $65^{\circ}C^{6}$. The obtained nanoemulsions were cooled to room temperature to crystallize the lipid and finally formed the active-loaded NLC.

The composition of coenzyme Q10 NLC system with different lipid ratio 70:30 can be seen in table 1. There prepared NLC was evaluated particle size and polydispersity index (PI) which were measured with Delsa NanoTM particle size analyzer. PI illustrates the variation on the sample. The small value of PI (<0.3) indicates that the sample is monodisperse⁷.

Table 1: Composition of coenzyme Q10 NLC system

Formulation	Coenzyme Q10 (%b/b)	Cetyl palmitate (%b/b)	Alpha tocopherol acetate (%b/b)	Tween 80 (%b/b)	Propyleneglycol (%b/b)
Coenzyme Q10	2.4	70	30	20	11

NLC: Nanostructured Lipid Carrier

Preparation of transdermal patches

Coenzyme Q10 NLC system with different lipid ratio 70:30 (cetyl palmitate and alpha tocopherol acetate) used as drug reservoir. HPMC 606 as rate controlling membrane was prepared by dissolving the polymer (10%, 15%, 20%) in aqua demineralized. Menthol was dissolved in ethanol were added into HPMC 606, cetostearyl alcohol was dissolved in ethanol were added into the mixture. The resultant dispersion was placed into circular patches diameter 3.5 cm and placed in desiccator for 24hour. The composition of coenzyme Q10 transdermal patches with can be seen in table 3.

Evaluation of transdermal patches

Thickness⁸

The thickness of patches was measured using Digital Vernier calipers at three different locations, and mean value was calculated.

Weight variation⁸

Weigt variation was studied by individually weighing three randomly selected patches. The determination was performed for each formulation and mean value was calculated.

Moisture content (MC)⁸

Moisture content was measured using weigher and stored patch in the desiccator for 24 hours.

Drug content

A prepared patch was added to 200-mL absolute ethanol and stirred vigorously for 2 hours. The contents were filtered,

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and Coenzyme Q10 was estimated spectrophotometrically at wavelength of 273 nm.

Drug homogeneity

To calculate drug drug homogenity, pacth equivalent a quarter part were accurately weighed, then was dissolved into ethanol and stirred for 2 hours. This resulting solution was then filtered through Whatmann filter paper then was determined by Spectrophotometric analysis at a wavelength of 273 nm.

Scanning Electron Microscopy (SEM)⁹

Surface morphology and shape of patch was studied using Scanning Electron Microscopy (SEM), Model Carl Zeiss MA10, USA at suitable magnification in room temperature. The microphotographs were observed for morphological characteristics and to confirm morphology of patch.

3. RESULT AND DISCUSSION

Evaluation of the prepared NLC

From the result of prepared NLC was evaluated particle size and polydispersity index which were measured with Delsa NanoTM particle size analyzer which were found to 153.766 \pm 1.220 nm and 0.198 \pm 0.014, can be seen in table 2.

Table 2: Evaluation particle size and polydispersity index

Formulation	Replicate	Evaluation		
rormulation		Particle size (nm)	Polydispersity index	
Coenzyme Q10 NLC (70:30)	1	153.50	0.214	
	2	155.10	0.187	
	3	152.70	0.194	
Mean ± SD		153.766 ± 1.220	0.198 ± 0.014	

From the results of particle size, it was found that all prepared coenzyme Q10 NLC have a particle size less than 0.2 µm, and as such are effective for transdermal applications. It was noticed that the small particle size impacted to the occlusivity increased due to surface of skin's contact area increased. Increasing the occlusive impacted to the hydration of stratum corneum which would be impacted to the drug released profile and flux^{10, 11}.Coenzyme Q10 NLC have a particle size less than 3 µm, and as such are effective for transdermal applications³. The Polydispersity Index is dimensionless and scaled such that values smaller than 0.05 are rarely seen other than with highly monodisperse standards. Values greater than 0.7 indicate that the sample has a very broad size distribution and is probably not suitable for the dynamic light scattering (DLS) technique¹².As shown in the table 2, polydispersity index meet the specification (PI < 0.3), so coenzyme Q10 NLC system lipid ratio 70:30 had monodisperse of particle size⁷.

Preparation of transdermal patches

Transdermal patch containing Coenzyme Q10 were prepared as per table 3.

Table 3:Composition of coenzyme Q10 transdermal patches

Formulation code	Coenzyme Q10 NLC (70:30)	HPMC 606 (10%)	HPMC 606 (15%)	HPMC 606 (20%)	Menthol	Cetostearyl alcohol
F1	1.604 mg	1 ml	-	-	1%	350 mg
F2	1.604 mg	-	1 ml	-	1%	350 mg
F3	1.604 mg	-	-	1 ml	1%	350 mg
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NLC: Nanostructured Lipid Carrier, HPMC: Hydroxy Propyl Methyl Cellulose

Organoleptic evaluation of transdermal patches

As shown in table 4, all of formulas indicated orange color, round shape, dry and slightly stiff, smooth surface texture, and menthol odor. Patch F1 and F2 have smooth surface better than F3 due to concentration of HPMC 606 as rate controlling membrane has more than other formulas, so there were more particles entrapped.

Table 4	l: Organ	oleptic of	oservation	of	coenzyme	Q10	transdermal
patches							

Formulation	Shape	Colour	Odor
code			
F1	Round, dry and slightly stiff,	Orange	Menthol
	smooth surface texture		
F2	Round, dry and slightly stiff	Orange	Menthol
	smooth surface texture	_	
F3	Round, dry and slightly stiff	Orange	Menthol
	rough surface texture		



Fig 1: Organoleptic of coenzyme Q10 transdermal patches. F1 (fig. A), F2 (fig. B), and F3 (fig. C).

Evaluation of transdermal patches

Thickness

This test was carried out to ensure the uniformity of thickness of each patch. The thickness was affected by the technique of pouring into the mold. Patch thickness was measured at 3 different points using the calipers. The results of the Coenzyme Q10 patch thickness test can be seen in table 5. The thickness range were $(1.92 \pm 0.003 \text{ to } 1.98 \pm 0.003 \text{ mm})$. This thickness value showed good characteristics of the patch¹³.

The results showed that the thickness F1 <F2 <F3, due to HPMC 606 has swelling so it would expand when dissolved to solvent. The higher the concentration of HPMC 606 increased, the thickness of the patch also increased. Thickness of the patch affected to the drug released and flux. Thickness increased impacted to the duration of release, the time of action more longer. Based on one way ANOVA test (0.068 > 0.05) there was no significant difference in the thickness value of the patch of each formula.

Table 5: Th	ickness test	of	coenzyme (Q10	transdermal	patches
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Formulation and	Thickness	of patch(Maan CD	
rormulation code	1	2	3	Wiean± SD
F1	1.90	1.90	1.95	1.92 ± 0.003
F2	1.95	1.95	2.00	1.97 ± 0.003
F3	2.00	2.00	1.95	1.98 ± 0.003

С

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Weight variation

Coenzyme Q10 patch with various concentration of HPMC 606 as the controlling membrane rate can be seen in table 6. If the weight of the patch decreases from the specified weight, then there was likely that one ingredient that has a reduced weight. If one of the reduced ingredients was the active ingredient, it would affect the amount of active ingredient in the penetration testing, therefore the weight was considered.

The test results showed F1 <F2 <F3, due to concentration of HPMC 606 increased, so the weight of the patch also increased. The weight range from 1.235 ± 0.006 to 1.422 ± 0.003 g. Based on the one way ANOVA test there were significant differences (0.00 < 0.05) in the weight parameter. Table 6: Weight variation test of coenzyme Q10 transdermal patches

Formulation code	Weight of	f patch (g)	Moon + SD	
	1	2	3	wiean ± 5D
F1	1.235	1.229	1.241	1.235±0.006
F2	1.315	1.307	1.309	1.310±0.004
F3	1.419	1.424	1.422	1.422±0.003

Moisture content (MC)

The moisture content (MC) range of 8.441 ± 0.077 for F3 to $9.514 \pm 0.073\%$ for F1, % MC value decreased as the HPMC 606 polymer increased. Low moisture content could maintain the stability of the preparation. MC requirement for patch preparation was<10%¹⁴. All formulas were meet the requirements.The moisture content result were shown in table 7.

Based on one way ANOVA test, there were significant differences in the moisture content value of each formulas (0.00 < 0.05). Prepared patch was expected low moisture content in order to increase stability and reduced wrinkles in storage for a long time¹⁵.

Table 7: Moisture content test of coenzyme Q10 transdermal patch	es
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Formulation and a	Moistur	e content	Marris CD	
rormulation code	1	2	3	mean ± SD
F1	10.040	9.927	9.659	9.875 ± 0.196
F2	9.430	9.564	9.549	9.514 ± 0.073
F3	8.457	8.357	8.509	8.441 ± 0.077

Drug content and Drug homogeneity

The drug content and drug homogeneity result were shown in table 8 and table 9. This test aim to ensure that the amount of coenzyme Q10 contained in the patch was at the right amount or in the specified range. Measurement of drug content in the patch matrix to obtain coenzyme Q10 levels using a optimun wavelength of 273 nm. The standard curve equation obtained is y = 0.00406x + 0.00797 with the coefficient of determination (R^2) of 0.99954. Measurement of drug content and drug homogeneity was carried out on the optimal formula with replication 3 times. The results of drug content ranged from 96.90% to 97.21% while drug homogeneity ranged from 97.18% to 98.03%. The calculation of patch homogeneity shows the % CV value is less than 2%, so it can be said that coenzyme Q10 is spread evenly on patch preparation¹⁵.Based on the results of the one way ANOVA test (0.889> 0.05 and 0.779 > 0.05) there were no significant differences of drug content and drug homogeneity of each formula.

Table 8: Drug content of coenzyme Q10 transdermal patches (data shows mean $[n{=}3]{\pm}SD)$

Evoluction	Formulations code						
Evaluation	F1	F2	F3				
Drug content (%)	97.21±1.90	97.20±0.53	96.90±0.92				
CV (%)	1.95	0.55	0.95				

Table 9: Drug homogeneity of coenzyme Q10 transdermal patches (data shows mean $[n{=}3]{\pm}SD)$

Evolution	Formulations code					
Evaluation	F1	F2	F3			
Drug homogeneity (%)	98.03±0.50	97.18±1.78	97.75±1.78			
CV (%)	0.51	1.83	1.82			

Scanning Electron Microscopy (SEM)

SEM photographs of the matrix patch was taken, surface morphology and drug distribution pattern of the transdermal patches sould be studied from SEM model Carl Zeiss MA10, USA with 56x magnification (Fig 2).

Scanning Electron Microscopy (SEM) images showed the upper surface of the patches in F1, F2 and F3. The greater of HPMC 606 concentration used, the greater the pore formed in the prepared patch, due to the swelling of HPMC 606 during Coenzyme Q10 patch preparation¹⁶. The pores on the patch are useful in the drug released from the matrix, the large pores allow the drug escape more easily. Penetration enhancement is an improving system that leads to the increase in the count of drugs through skin because of possessing different features like natural origin, favorable penetration enhancement and partitioning action in the skin by the oils¹⁷.





Fig 2: Photograph of Coenzyme Q10 patches surface with various levels of HPMC 606 using SEM with 56x magnification. F1 (fig. A), F2 (fig. B), dan F3 (fig. C).

4. CONCLUSION

Transdermal patch coenzyme Q10 preparations using NLC coenzyme Q10 as a drug reservoir and HPMC 606 (10%, 15%, and 20%) as the rate controlling membrane, all provide good characteristics. Based on the results of the one way ANOVA test(p-values > 0.05) there were no significant differences of evaluated for thickness, drug content and drug homogenity, except on the weight and moisture content of patch gives significant results.

5. REFERENCES

- Piao H, Mei O, Dengning X, Peng Q, Wenhua X, Yanzhi, S, Fude. 2011. In vitro – in vivo study of coq10-loaded lipid nanoparticles in comparison with nanocrystals. International Journal of Pharmaceutics 2011;419 (1-2):255-259.
- Mendes M, Nunus SCC, Sousa JJ, Pais AA, Vitorino C. Expanding transdermal delivery with lipid nanoparticles: a new drug-in-nlc-in-adhesive design. American Chemical Society 2017;14(6):2099–2115.
- Khurana S, Jain NK, Bedi PMS. Development and characterization of a novel controlled release drug delivery system based on nanostructured lipid carriers gel for meloxicam. Life Sciences 2013;93:763-772.
- Patel D, Dasguptas S, Dey S, Ramani YR, Ray S, Mazumder B. Nanostructured lipid carriers (nlc)-based gel for the topical delivery of aceclofenac:preparation, characterization, and in vivo evaluation. Sci Pharm 2012;80:749-764.
- Jaiswal P, Gidwabi B, Vyas A. Nanostructured lipid carriers and their curren application in targeted drug delivery. Artificial Cells, Nanomedicine, and Biotechnology 2014;44:27-40.
- Putranti AR, Primaharinastiti R, Hendradi E. Effectivity and physicochemical stability of nanostructured lipid carrier coenzyme q10 in different ratio of lipid cetyl palmitate and alpha tocopheryl acetate carrier. Asian Journal Of Pharmaceutical And Clinical Research 2017;10(2):146-152.
- Almeida MM, Bou-Chacra NA, Conte JD, Kaneko TM, Baby AR, Velasco MV. Evaluation of physical and chemical stability of nanostructured lipid carries

containing ursolic acid in cosmetic formulation. J Appl Pharm Sci 2013;3(01):005-8.

- 8. Parisvesh S, Sumeet D, Abhishek D. Design, evaluation, parameters and marketed products of transdermal patches. J Pharm Res 2010;3(2):235-240.
- Naseera K, Sajeeth CI, Santhi, K. Formulation, optimization and evaluation of matrix type of transdermal system of sisvastatin using permeation enhancers. International Journal of Current Pharmaceutical Research 2012;4(2):79-87.
- Brugè, Francesca, Damiani E, Puglia C, Offerta A, Armenia T. Nanostructured lipid carriers loaded with coq10: effect on human dermal fibroblasts under normal and uva-mediated oxidative conditions. International Journal of Pharmaceutics 2013;455:348-356.
- Teeranachaideekul V, Boonme P, Souto EB, Müller RH, Junyaprasert VB. Influence of oil content on physicochemical properties and skin distribution of Nile red-loaded NLC. J Control Release 2008;128(2):134-41.
- Gupta PV, Pandit JK, Kumar A, Swaroop P, Gupta S. Pharmaceutical nanotechnology novel nanoemulsionhigh energy emulsification preparation, evaluation and application. The Pharma Research 2010;3:117-138.
- Yener G, Uner M, Gonullu U, Yildirim S, Kilic P. Design of meloxicam and lornoxicam transdermal patches: preparation, physical characterization, ex vivo and in vivo studies. Cherm. Pharm. Bull 2010;58(11):1466-1473.
- Shinde AJ, Grala KC, More HN. Development and characterization of transdermal therapeutics system of tramadol hydrochloride. Asian Journal.Pharmaceutical 2008; 12(4):265-269.
- Duan XD, Chang JJ, Lin N. Formulation and development of dendrimer-based transdermal patches of meloxicam for the management of arthritis. Tropical Journal of Pharmaceutical Research 2015;14(4):583-590.
- Shirsand SB, Ladhane GM, Prathap S, Prakash PV. Design and evaluation of matrix transdermal patches of meloxicam. RGUHS Journal of Pharmaceutical Sciences 2012; 2(4):58-65.
- 17. A.C. Williams dan B.W. Barry. Penetration Enhancer. Advanced Drug Delivery Reviews 2004;56:603-618.

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