## **Original article**

# The Effect of Addition of Extracts of *Vernonia amygdalina* and *Moringa oleifera* in the Nutrition of Alloxan-Induced Diabetic Wistar Rats

Sokiprim Akoko<sup>1,\*</sup>, Benjamin Miaba Aleme<sup>2</sup>, Precious Ojo Uahomo<sup>3</sup>

<sup>1</sup>Department of Pharmacology, Faculty of Basic Clinical Sciences, College of Health Sciences, University of Port-Harcourt, Rivers State, Nigeria;

<sup>2</sup>Department of Biochemistry, Faculty of Sciences, University of Port-Harcourt, Rivers State, Nigeria; <sup>3</sup>Department of Biomedical Technology, School of Science Laboratory Technology, University of Port-Harcourt

<sup>3</sup>Department of Biomedical Technology, School of Science Laboratory Technology, University of Port-Harcourt, Rivers State, Nigeria.

## ARTICLE INFO:

Received: 06 July 2022 Accepted: 08 Aug 2022 Published: 31 Aug 2022

**Corresponding author \*** Akoko S, Department of Pharmacology, Faculty of Basic Clinical Sciences, College of Health Sciences

College of Health Sciences, University of Port-Harcourt, Rivers State, Nigeria. E-mail:

sokiprim.akoko@uniport.edu.ng

#### ABSTRACT:

The antidiabetic effect of co-administration of extracts of *Vernonia amydalina* and *Moringa oleifera* on alloxan-induced diabetic Wistar rats was evaluated. The study design involved 64 adult Wistar rats. They were randomly divided into eight different experimental groups of 8 animals each. The experiment lasted for 21 days for which animals were treated with extracts of of *Vernonia amydalina* and *Moringa oleifera*. Their body weight and blood glucose level were determined using weighing scale and hand held glucometer. The results showed that the co-administration of different combination doses of *V. amygdalina* and *M. oleifera* significantly (p<0.05) resulted to an increase in the body weight of experimental rats and significantly (p<0.05) reduced in a dose dependent manner the blood glucos in experimental rats when compare to diabetic control rats. The antidiabetic potentials of the co-administration of the leaf extracts of *V. amygdalina* and *M. oleifera* can be attribute to secondary constituents present in the plants which stimulates insulin production and release from the beta-cells, resulting in a fall in blood glucose to normal levels.

**Keywords:** Diabetes, *Vernonia amydalina, Moringa oleifera*, alloxan-induced, co-administration, Nutrition

## 1. INTRODUCTION

Diabetes mellitus is an endocrine system metabolic condition that causes complications with glucose, lipid, and protein homoeostasis [1]. The disease is present in all and is spreading quickly across the globe. However, the Kitava study, which examined the non-Westernized population of Kitava, one of the Trobriand Islands of Papua New Guinea, between 1989 and 1993, concluded that Kitava residents reportedly did not have high blood pressure, diabetes, obesity, ischemic heart disease, or stroke [2].

Absolute or relative impairments in insulin secretion and action, along with persistent hyperglycemia and disruptions of the metabolism of carbohydrates, lipids, and proteins, are characteristics of diabetes mellitus (DM) [3-5]. It is caused by a defect with the pancreas' ability to secrete insulin or by the cells' failure to use the insulin made by the beta-cells of the pancreatic islet of Langerhans [6, 7].

DM is a global burden that can lead to serious side effects like heart attack, stroke, and kidney failure. A person with diabetes mellitus must live with continual awareness of their condition, one or two daily insulin shots, and regular finger pricks to check their blood sugar levels, a restricted diet, and worry about complications. Even after taking into account enhanced surveillance, the prevalence of diabetes has more than doubled globally over the past three decades, greatly outpacing model estimates as diabetes today affects almost one in ten persons worldwide [8-10].

Over 86 percent of people in underdeveloped countries, according to the World Health Organization (WHO), rely on traditional remedies like herbs for their everyday requirements, and about 855 traditional medications contain crude plant extracts [8]. Sokiprim and colleagues showed that patients with type 2 DM received unintentionally suboptimal treatment due to the high cost of oral hypoglycaemic drugs. Additionally, because people in lower socioeconomic classes commonly choose alternative solutions, their health-seeking behaviors aren't necessarily consistent with conventional orthodox care. This is supported by confirmation from the World Health Organization, which encourages research into alternative diabetes treatments due to issues with insulin and the hazardous side effects of currently available synthetic oral glucose-lowering medications. Therefore, it is of utmost scientific interest to understand how food sources, natural antioxidants, and free radical scavengers contribute to the prevention and treatment of disease [11-14]. Over 150 million people worldwide have diabetes, making it one of the main causes of death [15]. Over 1.70 million Nigerians over the age of 15 have diabetes, and every year, about 70,000 children under the age of 15 are diagnosed with Type 1 diabetes. Obesity, population increase, and sedentary lifestyles are all contributing to the prevalence of diabetes, which is expected to reach over 360 million cases by 2030 [16].

In traditional medicine, more than 50% of plants are used to treat diseases that impact people, including diabetes, toothaches, diarrhea, dysentery, and skin infections. Numerous conventional medications have been developed from model compounds found in therapeutic plants. Over 400 traditional plant remedies for diabetes have been documented to date, but only a small number of these remedies have undergone scientific and medical testing to determine their usefulness [17]. The attributed hypoglycemic effects of these plants result from their capacity to restore pancreatic tissue function by increasing insulin secretion, inhibiting intestinal glucose absorption, or by facilitating metabolites in insulin-dependent processes. Therefore, using herbal medications for therapy has an impact on preserving beta-cells and reducing glucose fluctuation. The majority of these plants have been discovered to contain chemical elements, such as glycosides, alkaloids, phenols, terpenoids, and flavonoids that are frequently associated with having antidiabetic properties [18].

Tropical Africa is home to the plant *Vernonia amygdalina* [19]. It belongs to the Asteraceae family and is the source of a number of traditional medicines in the West African region [20-22]. It is frequently referred to as bitter leaf. The leaves are the plant parts that are most frequently used. The plant's young, succulent, and fresh leaves are typically recommended for laxative and treating conditions like diabetes, kidney function, malaria, fever, constipation, and high blood pressure [23].

The drumstick tree, *Moringa oleifera* (*M. oleifera*), is a member of the Moringaceae family. It is indigenous to the sub-Himalayan regions, although it is now widely grown and has naturalized in many tropical areas [24]. It is a fast-growing tree that is used in African folk medicine. Its production and administration are made simple by the ease with which it spreads by sexual and asexual mechanisms as well as the little requirement it has after being planted for soil nutrients and water [25]. It is an edible plant with significant medical and nutritional benefits that is used to heal a variety of illnesses, hence the term "wonder tree" [26]. Comprehensive research has been done on *M. oleifera* plant parts for the treatment of a variety of illnesses, including

typhoid fever, arthritis, malaria, swellings, skin conditions, parasitic diseases, hypertension, diabetes, liver disorders, and for boosting the immune system in patients with immunosuppressed individuals [27, 28]. Previous published scientific investigations on laboratory animals support the traditional usage of *M. oleifera* in treating diabetes [29]. *Moringa oleifera* extract has been linked to a wide range of pharmacological effects, including anticancer, anti-inflammatory, hypocholesterolemic, anti-atherosclerotic, antioxidant, neuro-protective, kidney-protective, and hepato-protective effects [27-33].

This study aimed to evaluate the effectiveness of combined extracts of *Vernonia amygdalina* and *Moringa oleifera*, two proven antidiabetic herbs, in the management of lipid problems often linked to chronic diabetes. According to Ugochukwu *et al.*, some minor food ingredients and secondary plant metabolites alter biological processes, which lower the risk of chronic diseases in people like diabetes mellitus. Understanding the manner of action of such agents may also require this [34]. Exploring and implementing newer approaches to attenuate DM is essential to reducing the burden from DM due to the high predictable rise in its burden, particularly in limited resource situations [35].

## 2. METHODOLOGY

#### 2.1. Preparation of Plant materials

Fresh leaves of Vernonia amygdalina Del. and Moringa oleifera Lam. were collected from a botanical garden at Aluu Community in the Obio/Akpor Local Government Area of Rivers State, Nigeria, and identified in the herbarium of the Department of Plant Science and Biotechnology in the University of Port Harcourt. To get rid of dust and dirt, the leaves were rinsed multiple times with clean tap water and given time to thoroughly drain. A knife was used to separate and chop the plant components, and then one kilogram (1kg) of each of V. amygdalina and M. oleifera was homogenized in 1.95 and 2.25 liters of ethanol that was 80% (v/v) by volume. The mixtures were placed in the refrigerator at 4°C for 48 hours to fully extract the active ingredients from the plants. These were first filtered using cheesecloth and then Whatman No. 1 filter paper, and the filtrates were then concentrated using a rotary evaporator in a vacuum at a low temperature  $(37-40^{\circ}C)$  to about one tenth the original volume. For V. amygdalina and M. oleifera, the concentrates produced 41.05g (4.105 percent) and 35.21g (3.521 percent) of greenish/greenish-brown oily substances when left uncovered in a water bath at 40°C for complete drying. The extracts were then kept in a refrigerator at a temperature of 2 to 8°C until usage.

#### 2.2. Toxicity studies

Acute toxicity employed for *Vernonia amygdalina* was the method by Lorke [36] as described by Okoli *et al.* [9] while the acute toxicity and  $LD_{50}$  of *Moringa oleifera* ethanolic leaves extract in rats was estimated using the method as described by Osman *et al.* [37].

## 2.3. Phytochemical Screening of Vernonia amygdalina and Moringa oleifera leaves

Alkaloids, tannins, saponnins, flavonoids, phenol, Terpenes, steroid and anthraquinones were quantitatively determined using standard methods [38-41].

#### 2.4. Procurement of Animal

For this investigation, adult Wistar rats of either sex weighing 140–210g were used. They were acquired from the Animal House of the Department of Pharmacology at the University of Port Harcourt in River State, Nigeria, and were acclimatized for two weeks. They were kept in a conventional laboratory environment with  $28^{\circ}$ C temperature ( $28\pm2^{\circ}$ C), relative humidity ( $46\pm6\%$ ), a 12-hour light/dark cycle, and adequate ventilation. The animals were given access to water and a commercial feed (Vital Feed Nig. Ltd.) *ad libitum*. Twelve hours prior to the experiments, food was withheld, although water was always available for free.

#### 2.5. Ethical Clearance

According to the recommendations made by the University of Port Harcourt's Research Ethics Committee, all methods used in this study were carried out in compliance with the fundamental principles of animal-based research.

## 2.6. Drug Purchase and Preparation

Glibenclamide (GBC) was obtained from E-Blend Pharmacy, a licensed pharmacy located inside the University of Port Harcourt. In order to prepare the powder for administration to the test animals, the tablets were crushed into a fine powder and the proper concentrations produced in distilled water. Alloxan monohydrate, another substance utilized, was also bought from the same pharmacy to cause diabetes in rats.

#### 2.7. Induction of Diabetes

Alloxan monohydrate, freshly made with distilled water as the vehicle, was diluted to a concentration of 150mg/kg body weight and administered intraperitoneally to rats to cause diabetes. Three days later, diabetes was identified in alloxaninduced rats with Random Blood Glucose (RBG) levels

200mg/dL. Glucose levels were monitored using a hand held glucometer (Accu-CHEK) to test blood samples taken from the tail vein.

## 2.8. Experimental Design

Sixty-four (64) rats were divided into eight different experimental groups of 8 animals each. Group 1 (normal control group) received 0.5ml dimethylsulfoxide (DMSO), group 2 (diabetic control group) received placebo, 0.5ml dimethylsulfoxide (DMSO), group 3 (glibenclamide group) received 0.2mg/kg body weight glibenclamide (GBC, standard drug) orally, group 4 (*V. amygdalina* group) received 200mg/kg body weight of *V. amygdalina* extract, group 5 (*M. oleifera* group)received 200mg/kg body weight of *N. oleifera* extract, group 6 (low dose combination of *V. amygdalina* and *M. oleifera* extracts, group 7 (medium dose combination of *V. amygdalina* and *M. oleifera* extracts, group 7 (medium dose combination of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts, group 7 (medium dose combination of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* and *M. oleifera* extracts) received 200mg/kg each of *V. amygdalina* extracts) received 200mg/kg each of *V. amygdalina* extracts) received 200mg/kg each of *V. amygdalina* extrac

*oleifera* extracts and group 8 (high dose combination of *V. amygdalina* and *M. oleifera* extracts) received 300mg/kg each of *V. amygdalina* and *M. oleifera* extracts. Upon administration, experimental rats were allowed access to feed and water.

Table 1: Experimental design					
Group	No. of Rate	s Treatment			
Group 1	8	Normal Control			
Group 2	8	Alloxan Control			
Group 3	8	0.2mg/kg Glibenclamide (GBC)			
Group 4	8	200mg/kg V. amygdalinaExtract			
Group 5	8	200mg/kg M. oleiferaExtract			
Group 6	8	100mg/kg V. amygdalina + 100mg/kg M. oleiferaExtracts			
Group 7	8	200mg/kg V. amygdalina + 200mg/kg M. oleiferaExtracts			
Group 8	8	300mg/kg V. amygdalina + 300mg/kg M. oleiferaExtracts			

The plant extracts were administered orally. The dosage of the extract was determined from preliminary studies in the laboratory. The duration of treatment was 21 days after induction of diabetes.

#### 2.9. Body weight and Blood glucose determination

Body weight and blood glucose levels were monitored in order to assess the combined effects of *V. amygdalina* and *M. oleifera* extracts in treated diabetic rats. The blood glucose level was monitored using a glucometer (Accu-CHEK) to measure the blood glucose level from the tail vein using a tail clip. Body weight of the animals was evaluated using a laboratory weighing scale. Experimental animals had their body weight and blood glucose levels checked before starting therapy, on days 7, 14, and 21, and also before and after inducing diabetes.

## 2.10. Method of Data Analysis

Using the statistical program SPSS for Windows version 23.0, descriptive statistics were used to examine the data from this investigation and provided as mean standard error of mean of three determinations (mean  $\pm$  SEM) (SPSS Inc. Chicago IL). Multiple comparison tests and analysis of variance (ANOVA) were used to differentiate differences between means. P values less than 0.05 were considered significant. The mean  $\pm$  SEM body weight and blood glucose values were given in g and mg/dL respectively.

#### 3. RESULTS

*V. amygdalina* and *Moringa oleifera* leaves both have oral LD50 values greater than 3000 mg/kg and 6500mg/kg body weight, respectively, according to an acute toxicity test [37,42]. *V. amygdalina* and *Moringa oleifera* leaves were examined for their phytochemical composition, which included alkaloids, tannins, saponins, flavonoids, phenol, terpenes, and steroids (Table 3). Table 2 displays the percentage yield of the ethanolic leaf extracts of *V. amygdalina* and *M. oleifera*.

Table 2: Percentage yield of Plant leaves extracts				
Plant Yield (%)		Yield (g)		
V. amygdalina	4.105	41.05		
M. oleifera	3.521	35.21		

#### Table 3: Phytochemical (quantitative) analysis of the plants

Metabolites	Test	V. amygdalina	M. oleifera
Alkaloids	a. Mayer's test	+	+
	b. Dragendorf's test	+	+
	c. Wagners reagent	+	+
Tannins	a. Lead Sub-acetate	+	+
	b. Ferric chloride test	+	+
Saponins	a. Frothing test	+	+
Flavonoids	a. Ferric chloride test	+	+
	b. NaOH test	+	+
	c. Shinoda test	+	+
Phenols	a. Ferric chloride test	+	+
Terpenes	a. Lieberman-Buchard	+	+
	b. Salkowski test	+	+
Sterols	a. Lieberman-Buchard	+	+
	b. Salkowski test	+	+
Anthraquinones	a. Borntrager's test	-	•

**Key:** + presence of constituent, absence of constituent

 Table 4: Effect of Vernonia amygdalina and Moringa oleifera extract

 combination on the body weight of diabetic rats

 Body Weight (a)

bouy weight (g)				
Group	Pre-	Day 7	Day 14	Day 21
	Treatment			
Normal Control	$165.86 \pm 4.20$	170.57±2.66	174.66±4.83	181.23±5.24
Alloxan Control	151.96±13.0	$142.20 \pm 8.71 *$	136.54±7.63*	118.62±9.12*
0.2mg/kg	$152.56 \pm 0.61$	168.61±9.31*	174.24±7.48*#	171.40±10.22
Glibenclamide		#		
(GBC)				
200mg/kg VAE	$153.52 \pm 5.74$	$170.45 \pm 8.70*$	177.10±6.12*#	$175.47{\pm}12.21$
		#		*#
200mg/kg MOE	$154.15\pm6.05$	$168.65 \pm 4.95 *$	176.88±7.65*#	$173.15{\pm}11.62$
		#		*#
100mg/kg VAE +	$151.30\pm6.17$	$171.25 \pm 6.85 *$	175.78±9.68*#	172.61±11.25
100mg/kg MOE		#		*#
200mg/kg VAE +	153.32±11.18	$174.40{\pm}10.05$	178.85±6.10*#	171.43±7.44#
200mg/kg MOE		*#		
300mg/kg VAE +	154.30±10.13	$175.40{\pm}10.15$	180.28±7.23*#	172.54±6.57*
300mg/kg MOE		*#		#
Values are repres	sented in mea	n+SFM valu	es marked wi	th (*) differ

Values are represented in mean $\pm$ SEM, values marked with (\*) differ significantly from normal control value (\*p 0.05) while those marked with (#) differ significantly from Alloxan control group (#p 0.05). VAE = *Vernonia amygdalina* Extract, MOE = *Moringa oleifera* Extract



Fig 1: Bar Chart showing the effect of *Vernonia amygdalina* and *Moringa oleifera* extract combination on the body weight of diabetic rats

 Table 5: Effect of Vernonia amygdalina and Moringa oleifera extract

 combination on blood glucose level of diabetic rats

 Blood Glucose Level(mg/dL)

Group	Pre-	Day 7	Day 14	Day 21
	Treatment			
Normal Control	60.5±2.14	62.2±2.21	62.8±1.84	61.1±2.18
Alloxan Control	255.6±6.31	268.4±7.16*	$282.2\pm6.08*$	310.5±12.23*
0.2mg/kg	$248.4 \pm 7.28$	196.5±6.82*	#200.4±8.12*	#178.0±7.26*#
Glibenclamide (GBC	C)			
200mg/kg VAE	238.6±6.18	241.7±7.30*	#220.2±8.24*	#198.5±5.84*#
200mg/kg MOE	235.9±6.08	238.5±6.71*	#217.8±5.12*	#196.2±4.92*#
100mg/kg VAE +	237.8±6.33	217.5±6.81*	#195.8±7.10*	#188.1±7.42*#
100mg/kg MOE				
200mg/kg VAE +	248.9±7.22	202.4±8.47*	#190.5±5.11*	#181.7±6.73*#
200mg/kg MOE				
300mg/kg VAE +	236.8±6.88	200.5±7.21*	#189.2±8.05*	#180.4±6.46*#
300mg/kg MOE				

Values are represented in mean $\pm$ SEM, values marked with (\*) differ significantly from normal control value (\*p 0.05) while those marked with (#) differ significantly from Alloxan control group (#p 0.05). VAE = *Vernonia amygdalina* Extract, MOE = *Moringa oleifera* Extract





#### 4. DISCUSSION

Numerous plant treatments have been utilized in traditional medicine for a number of ailments, including the treatment of diabetes mellitus [43]. Scientific research has proven that several of these herbal remedies have biological effects that counter diabetes mellitus and its consequences [44]. The biochemicals found in the plant components have been credited for some of these plants' therapeutic qualities. Nwaoguikpe [45], Adikwu et al. [46] and Asante et al. [47] investigated the effects of V. amygdalina on diabetic rat models and came to the conclusion that it has an antidiabetic impact. In a similar vein, a study by Onyagbodor and Aprioku [48] confirmed the anti-diabetic activity of Moringa oleifera, while a review by Fatoumata et al. [49] provided comprehensive diabetes research employing Moringa oleifera that demonstrated the impact of the Moringa plant in diabetic rat models.

The goal of the current investigation is to determine whether co-administering *Vernonia amygdalina* and *Moringa oleifera* extracts in the nutrition of Wistar rats will have an anti-diabetic impact. Both plants contained alkaloids, tannins, saponins, flavonoids, phenol, terpenes, and steroids, according to the phytochemical analysis. These phytochemicals help to explain some of the effects of using these plants as herbal remedies. Saponins [50,51], flavonoids [52-54], alkaloids [55,56], steroids and terpenes [57,58] have been reported to have good antidiabetic, hypolipidemic and antihyperglycemic activities [59-62].

Alloxan, a well-known diabetogenic chemical and cytotoxic glucose analogue utilized in diabetes research, was used to cause diabetes in Wistar rats. Alloxan has two distinct pathological effects. First, by specifically inhibiting glucokinase, the beta cell's glucose sensor, it inhibits glucose-induced insulin secretion. Second, by causing the formation of reactive oxygen species (ROS), which causes the selective necrosis of beta cells, it results in an insulindependent diabetes state. These two actions can be attributed to the particular chemical characteristics of alloxan, with the beta cell's selective uptake and accumulation of alloxan serving as the common denominator. This led to pancreatic malfunction and the bulk of the islets of Langerhans' -cells being destroyed. Untreated diabetic animals lost weight overall and their blood glucose levels rose as a result of pancreatic dysfunction brought on by the disease. The action of the alloxan results in the necrosis-induced death of betacells [24].

The findings of this study revealed that untreated diabetic rats (alloxan control) significantly lost weight when compared to non-diabetic rats (normal control), whereas experimental rats' body weights significantly increased after receiving doses of 200mg/kg of V. amygdalina and M. oleifera, respectively. The co-administration of different combination doses of V. amygdalina and M. oleifera also significantly resulted to an increase in the body weight of experimental rats when compared with non-diabetic (normal control) rats and untreated diabetic rats (alloxan control) as shown in Table 4 and Figure 1. Additionally, when compared to normal and alloxan control rats, experimental rats treated with gilbenclamide had a rise in body weight. Therefore, experimental rats gained weight when V. amvgdalina and M. oleifera were administered together. This supports the conclusions reached by Minari [63] and Efiong et al. [64].

Fasting animals' blood glucose levels provide information on the amount of insulin in the bloodstream and the sensitivity of peripheral tissues to insulin's effects. Wistar rats' blood glucose levels are known to range from 50 to 135mg/dL, and readings above 200 mg/dL are typically indicative of severe hyperglycemia and the development of diabetes [65].

In this investigation, the control group consisted of water and dimethyl sulfoxide (DMSO). According to the findings, alloxan significantly raised blood glucose levels as compared to control rats, although blood glucose levels in control rats were within the previously mentioned normal range. Throughout the course of the experiment, the rats given alloxan showed a continuous rise in blood glucose levels (hyperglycemia). This is in line with other research that found that alloxan causes permanent diabetic mellitus 24 hours after administration [24,48,65].

Additionally, it was found that the elevated blood glucose levels of diabetic rats were decreased by individual doses of V. amygdalina and M. oleifera as well as by coadministration of V. amygdalina and M. oleifera extracts in a dose-dependent manner as shown in Table 5 and Figure 2. These effects were seen in comparison to non-diabetic rats (normal control) over time. Glibenclamide, a common medication, dramatically lowered the blood glucose level in diabetic rats. Gilbenclamide is a sulfonylurea that lowers blood sugar by encouraging pancreatic beta-cells to produce more insulin and by boosting the accumulation of glycogen in the liver. However, it may not be effective in alloxaninduced diabetic animals because alloxan treatment permanently destroys the beta cells [66]. As with the coadministered extracts, it also reduced the hyperglycemia brought on by alloxan. Co-administration of the extracts outperformed individual dose administration of V. amygdalina and M. oleifera extracts in terms of effectiveness. Therefore, experimental rats may develop normoglycemia if V. amygdalina and M. oleifera are administered together. The findings by Efiong et al. [64], Asante et al. [47], Ebong et al. [67] and Onyagbodor and Aprioku [48] are in agreement with this. The coadministered extract's antihyperglycemic efficacy may have been influenced by auxiliary components. This antihyperglycemic effect may be brought on by the terpenes and other bitter components found in these plants, which may also stimulate insulin production and release from beta cells. The findings imply that V. amygdalina and M. oleifera may play a dose-dependent protective role on pancreatic beta-cells and/or may have an insulin-like impact on boosting tissues by either peripheral glucose uptake/utilization or by blocking hepatic gluconeogenesis [68].

There is a chance of amplifying the insulin-like phytochemicals in these two extracts by combining them since the impact is similar to that of gilbenclamide, according to the evaluation of the antidiabetic effects demonstrated by the co-administration of V. amygdalina and M. oleifera. Some phytochemicals in anti-diabetic plants are known to have characteristics that mimic and function somewhat like insulin [69]. Therefore, it is possible that coadministration of these plants will stimulate insulin receptors, aid in the uptake and metabolism of glucose, and improve insulin sensitivity. Therefore, a combination of V. amygdalina and M. oleifera leaf extracts may be more advantageous and useful in managing diabetes as an antioxidant defense system, providing a comprehensive repertoire of free radical quenching components, and preventing atherosclerosis in diabetes mellitus.

## 5. CONCLUSION

Due to a potential synergy between the bioactive secondary chemicals from these two plants, the administration of mixed extracts from both plants' leaves demonstrated a potential impact. Because of this, it seems as though the leaves of these plants have a complement of bioactive substances, which could explain why they have a hypoglycemic effect. The utility of the combination herbal remedy as an anti-diabetic drug has been confirmed after Wistar rats that had been given alloxan-induced diabetes showed that it had an anti-diabetic effect. The current research backs up the traditional use of *M. oleifera* and *V. amygdalina* in diabetes control with scientific data.

## 6. REFERENCES

- Van den Berghe G, Wilmer A, Hermans G, Meersseman W, et al. Intensive insulin therapy in the medical ICU. N Engl J Med. 2006; 354:449-61.
- Liindeberg S and Lundh B. Apparent absence of stroke and ischaemic heart disease in a traditional Melanasian island: a clinical study in Kitava. J Intern Med. 1993; 233:269-75
- 3. Adeneye AA, Benebo AS. Pharmacological evaluation of a Nigerian polyherbal health tonic tea in rat. African Journal of Biomedical Research. 2007; 10: 249-55.
- Agrawal RP, Sharma P, Pal M, Kochar A, Kochar DK. Magnitude of dyslipedemia and its association with micro and macro vascular complications in type 2 diabetes: a hospital based study from Bikaner (Northwest India). Diabetes Research and Clinical Practice. 2006; 73:211-4.
- Scott AI, Clarke BE, Healy H, D'Emden M, Bell SC. Microvascular complications in cystic fibrosis-related diabetes mellitus: a case report. JOP. Journal of the Pancreas, 2000; 1:208-10.
- Gordon L, Ragoobirsingh D, St Errol YA, Choo-Kang E, McGrowder D, Martorell E, *et al.* Lipid profile of type 2 diabetic and hypertensive patients in the Jamaican population. Journal of Laboratory Physicians. 2010; 2: 25-30.
- 7. Marles RJ, Farnsworth NR. Antidiabetic plants and their active constituents. Phytomedicine. 1995; 2:137-89.
- Nimenibo-Uadia R. Effect of aqueous extract of *Canavalia ensiformis* seeds on hyperlipidaemia and hyperketonaemia in alloxan-induced diabetic rats. Biokemistri. 2003; 15:7-15.
- Okoli CO, Ibiam AF, Ezike AC, Akah PA, Okoye TC. Evaluation of antidiabetic potentials of Phyllanthusniruri in alloxan diabetic rats. African Journal of Biotechnology. 2010; 9: 248-59.
- Owulade MO, Eghianruwa KI, Daramola FO. Effects of aqueous extracts of Hibiscus sabdariffa calyces and *ocimun gratissimum* leaves on interstinal transit in rats. African Journal of Biomedical Research. 2004; 7:31-4.

- Afieroho OE, Ollonrnwi KV, Elechi N, Okwubie L, Okoroafor D, and Abo KA. Free radical scavenging potentials of and levels of some heavy metals in *Plerotus flabellatus* Berk and Broome (Pleurotaceae). The Global Journal of Pharmaceutical Research 2013; 2: 1807-12
- Chinenye S, Onyemelukwe GC, Johnson TO, Oputa RN, Oluwasanu M. Diabetes Advocacy and Care in Nigeria. Port Harcourt, Nigeria: Diabetes Association of Nigeria; 2014.
- Jaja P, Akoko S, Bestman A, & Iragunima A. Healthseeking behavior of Port Harcourt City Residents: A Univariate Comparison between the Upper and Lower Socio Economic Classes. The Nigerian Health Journal, 2016; 15: 141-22.
- 14. Sokiprim A, Dagogo MO, and Roseline MA. Medication Adherence and its Determinants among Patients with Type-2 Diabetes Attending University of Port Harcourt Teaching Hospital. World Journal of Advance Healthcare Research, 2022; 6: 62-71
- 15. Oyedepo TA, Babarinde SO, Ajayeoba TA. Evaluation of anti-hyperlipidemic effect of aqueous leaves extract of Moringaoleifera in alloxan induced diabetic rats. International Journal of Biochemistry Research & Review. 2013 Jul 1; 3:162-70.
- Rang HP, Dale MM, Ritter JM, Moore PK. Rang and Dale's Pharmacology. 5th Edition, Churchill Livingstone London. 2005; pp: 385-88.
- 17. Rockwood JL, Anderson BG, Casamatta DA. Potential uses of *Moringa oleifera* and an examination of antibiotic efficacy conferred by *M. oleifera* seed and leaf extracts using crude extraction techniques available to underserved indigenous populations. International Journal of Phytotherapy Research. 2013; 3:61-71.
- Rotimi SO, Omotosho OE, Rotimi OA. Persistence of acidosis in alloxan-induced diabetic rats treated with the juice of *Asystasia gangetica* leaves. Pharmacognosy Magazine. 2011; 7(25):25-30.
- Areghore EM, Makkar HPS, and Becker K. "Chemical composition and tannins in leaves of some browse plants from Delta (Central Nigeria) eaten by ruminants," *Proceedings of the Society of Nutrition Physiology*, 1997; vol. 5, pp. 11–1.
- 20. Akah PA, Okafor CI. Blood sugar lowering effect of *Vernonia amygdalina* (Del) in an experimental rabbit model. Phytother. Res. 1992; 6:171-3.
- Alebachew M, Kinfu Y, Makonnen E, BekuretsionY, Urga K. and Afework M. "Toxicological evaluation of methanol leaves extract of *Vernonia bipontiniVatke* in blood, liver and kidney tissues of mice," *African Health Sciences*, 2014; 14(4): 1012–24.
- 22. Amole OO, Izegbu MC, Onakoya JAA. and Dada MO. "Toxicity studies of the aqueous extract of *Vernonia amygdalina*," *Biomedical Research*, 2006; 17: 39–40.

- Leung KK, Leung PS. Effects of hyperglycemia on angiotensin II receptor type 1 expression and insulin secretion in an INS-1E pancreatic beta-cell line. Jop. 2008; 9: 290-9.
- Szkudelski T. The mechanism of alloxan and streptozotocin action in B cells of the rat pancreas. Physiological Research. 2001; 50: 536-46.
- Tasaduq SA, Singh K, Sethi S, Sharma SC, Bedi KL, Singh J, *et al.* Hepatocurative and antioxidant profile of HP-1, a polyherbal phytomedicine. Human & Experimental Toxicology. 2003; 22:639-45.
- Gopalakrishnan L, Doriya K, Kumar DS. Moringaoleifera: A review on nutritive importance and its medicinal application. Food Sci Hum Wellness 2016; 5:49–56.
- 27. Akoko S, Siminialayi IM. and Obianime AW. Evaluating the Effect of *Moringa* (K Formula Dietary Supplement) On Renal Function among HIV Positive Patients on TDF Regimen: A Longitudinal Study of Nigerians. International Journal of Public Heath, Pharmacy and Pharmacology, 2017; 2: 18-32
- Kou X, Li B, Olayanju JB, Drake JM, Chen N. Nutraceutical or pharmacological potential of Moringaoleifera Lam. Nutrients 2018; 10: 343.
- Yassa HD, Tohamy AF. Extract of *Moringa oleifera* leaves ameliorates streptozotocin-induced diabetes mellitus in adult rats. ActaHistochem. 2014; 116: 844-54.
- 30. Peixoto JRO, Silva GC, Costa RA, de Sousa Fontelle, JR, Veira GH, et al. *In vitro* antibacterial effect of aqueous and ethanolic Moringa leaf extract. Asian Pacific Journal of Tropical Medicine 2011, 4: 201-4.
- 31. Saalu LC, Ogunlade B, Ajayi GO, Oyewopo AO, Akunna GG. and Ogunmodede OS. The hepatoprotective potentials of *Moringa oleifera* leaf extract on alcohol induced hepato-toxicity in Wistar rat. American Journal of Biotechnology and Molecular Sciences, 2012; 2: 6-14.
- 32. El-bakry K, Toson E, Serag M. and Aboser M. Hepaprotective effect of *Moringa oleifera* leaves extract against carbon tetrachloride induced liver damage in rats. World Journal of Pharmacy and Pharmaceutical Sciences 2016; 5(5): 76-89.
- 33. Omeodu SI, Aleme BM, Akoko S, Uahomo PO. Hepato-ameliorative effect of aqueous extract of *Moringa oleifera* stem bark on paracetamol-induced liver injury in wistar rats. Gal Int J Health Sci Res. 2022; 7(2): 46-54
- 34. Ugochukwu N H, Babady N E, Cobourne M and Gasset S R. The effect of gongronemalatifolium extracts on serum lipid profile and oxidative stress indices in hepatocytes of diabetic rats. J Biosci. 2003; 28: 1 – 5
- 35. Siminialayi I.M., Emem-Chioma P.C. Type 2 diabetes mellitus: A review of pharmacological treatment. Nigerian Journal of Medicine: Journal of the National

Association of Resident Doctors of Nigeria 2006; 3: 207-14.

- 36. Lorke D. A New Approach to Practical Acute Toxicity Testing. Archives of Toxicology 1983; 53: 275-87.
- 37. Osman HM, Shayoub ME, Babiker EM, Faiza AO, Munzir ME Ahmed, Bashier Osman, et al. Assessment of acute toxicity and LD50 of *Moringa oleifera* ethanolic leave extract in albino rats and rabbits. Journal of Medical and Biological Science Research 2015; 1(4): 38-43
- Harborne JB. Phytochemical methods: A guide to modern techniques of plant analysis. Chapman and Hall Ltd, London. 1973; Pp. 279.
- Van-Burden T P, Robinson W C. Formation of complexes between protein and tannin acid. Journal of Agriculture and Food Chemistry. 1981;1:77–82
- 40. Obadoni BO and Ochuko PO. Phytochemical Studies and Comparative Efficacy of the Crude Extracts of Some Homeostatic Plants in Edo and Delta States of Nigeria. Global Journal of Pure and Applied Science, 2001; 8: 203-8.
- Uahomo PO, Isirima JC, Akoko S. Evaluation of Phytochemicals and Bioactive Properties in Leaf and Root Parts of *Cyathula prostrata* (Pasture Weed) – A Qualitative and Quantitative Analysis. Asian Plant Research Journal 2022; 9: 8-16.
- 42. Olanrewaju CA, Idris HS, Okwute SK, Olayanju S. Phytochemical analysis and acute oral toxicity of aqueous extracts of *Vernonia amygdalina* (Delile) and *Nauclea latifolia* (Smith). International Journal of Current Research 2015; 7: 11269-73.
- Sofowora A. Medicinal plants and traditional medicine in Africa, Reprinted ed. Ibadan: Spectrum book Ltd., 2006; pp. 150-153.
- 44. Ojiako OA, Nwanjo HUI. *Vernonia amygdalina* hepatotoxic or hepatoprotective? Response from biochemical and toxicity studies in rats. Afr J Biotechnol. 2006; 5:1648-51.
- Nwaoguikpe Reginald Nwazue. The effect of extract of bitter leaf (*Vernonia amygdalina*) on blood glucose levels of diabetic rats. Int J Biol Chem Sci 2010; 4: 721-9.
- 46. Adikwu U. Michael, B. Uzuegbu David, C. Okoye Theophine, F. Uzor Philip, et al. Antidiabetic effect of combined aqueous leaf extract of *Vernonia amygdalina* and metformin in rats. Journal of Basic and Clinical Pharmacy 2010; 1: 197-202.
- 47. Asante Du-Bois, Emmanuel Effah-Yeboah,1 Precious Barnes, Heckel Amoabeng Abban, Elvis Ofori Ameyaw, Johnson Nyarko Boampong, Eric Gyamerah Ofori and Joseph Budu Dadzie. Antidiabetic Effect of Young and Old Ethanolic Leaf Extracts of Vernonia amygdalina: A Comparative Study. Journal of Diabetes Research 2016; Article ID 8252741.

- Onyagbodor Olubunmi Adebambo and Aprioku Jonah Sydney. *Moringa oleifera* leaf extract inhibits diabetogenic effect of alloxan in rats. IOSR Journal Of Pharmacy 2017; 7(10): 07-12
- 49. Fatoumata BA., Mamadou Saïdou BAH., Mohamet Sene, Joseph Koulanzo Sambou, Modou Mbacké Gueye and El Hadji Makhtar BA. Antidiabetic properties of *Moringa oleifera:* A review of the literature. Journal of Diabetes and Endocrinology 2020; 11: 18-29
- Zheng T, Shu G, Yang Z, Mo S, Zhao Y, Mei Z. Antidiabetic effect of total saponins from *Entada phaseoloides* (L.) Merr. in type 2 diabetic rats. J Ethnopharmacol. 2012; 139: 814-21.
- 51. Elekofehinti OO, Kamdem JP, Kade IJ, Rocha JBT, Adanlawo IG. Hypoglycemic, antiperoxidative and antihyperlipidemic effects of saponins from *Solanum anguivi* Lam. fruits in alloxan-induced diabetic rats. South African J Bot. 2013; 88: 56-61.
- 52. Okokon JE, Ita BN, Udokpoh AE. The in-vivo antimalarial activities of *Uvaria chamae* and *Hippocratea africana*. Ann Trop Med Parasitol. 2006; 100: 585-90.
- 53. Brahmachari G. Bio-flavonoids with promising antidiabetic potentials: A critical survey Opportunity. Challenge and scope of Natural products in Medicinal chemistry 2011; 25:187-212.
- 54. Zheng XK, Zhang L, Wang WW, Wu YY, Zhang QB, Feng WS. Anti-diabetic activity and potential mechanism of total flavonoids of *Selaginella tamariscina* (Beauv.) Spring in rats induced by high fat diet and low dose STZ. J Ethnopharmacol. 2011; 137: 662-8.
- 55. Agrawal R, Sethiya NK, Mishra SH. Antidiabetic activity of alkaloids of *Aerval anata* roots on streptozotocin-nicotinamide induced type-II diabetes in rats. Pharm Biol. 2013; 51: 635-42.
- Tiong SH, Looi CY, Hazni H, Arya A, Paydar M, Wong WF, et al. Antidiabetic and antioxidant properties of alkaloids from *Catharanthus roseus* (L.) G Don. Molecules 2013; 18: 9770-84.
- 57. Kunyanga CN, Imungi JK, Okoth M, Momanyi C, Biesalski HK, Vadivel V. Antioxidant and antidiabetic properties of condensed tannins in acetonic extract of selected raw and processed indigenous food ingredients from Kenya. J Food Sci. 2011; 76: C560-7.
- Daisy P, Jasmine R, Ignacimuthu S, Murugan E. A novel steroid from *Elephantopus scaber* L. an ethnomedicinal plant with antidiabetic activity. Phytomedicine 2009; 16: 252-7.
- 59. De Tommasi N, De Simone F, Cirino G, Cicala C and Pizza C. Hypoglycemic effects of sesquiterpene glycosides and polyhydroxylated triterpenoids of *Eriobotrya japonica*. Planta Med. 1991; 57: 414-6.
- 60. Reher G, Slijepcevic M, Krans L. Hypoglycaemic activity of triterpenes and tannins from *Sarcopterium*

spinosum and two sanguisorba species. Planta Med. 1991; 57:57-8.

- Zarzuelo A, Jimenez I, Gamez MJ, Utrilla P, Fernadez I, Torres MI, et al. Effects of luteolin 5-O-beta-rutinoside in streptozotocin-induced diabetic rats. Life Sci. 1996; 58: 2311-6.
- 62. Yamabe N, Kang KS, Matsuo Y, Tanaka T, Yokozawa T. Identification of antidiabetic effect of iridoid glycosides and low molecular weight polyphenol fractions of *CorniFructus*, a constituent of Hachimi-jiogan, in streptozotocin-induced diabetic rats. Biol Pharm Bull. 2007; 30: 1289-96.
- 63. Minari JB. Hepatoprotective effect of methanolic extract of *Vernonia amygdalina* leaf. J Nat Prod. 2012; 5(2012):188-92.
- 64. Efiong EE, Igile GO, Mgbeje BIA, Otu EA and Ebong PE. Hepatoprotective and anti-diabetic effect of combined extracts of *Moringa oleifera* and *Vernonia amygdalina* in streptozotocin-induced diabetic albino Wistar rats. Journal of Diabetes and Endocrinology 2013; 4: 45-50
- 65. Carvalho, E.N., Carvalho, N.A.S. and Ferreira, L.M. Experimental model of induction of *Diabetes mellitus* in rats. Acta Cirurgica Brasileira 2003; 18: 1-5.
- Larner J, Haynes C. The Pharmacological basis of therapeutics, 5th ed, New York: Macmillan Publishing Co. Insulin and Hypoglycaemic Drugs, Glycogen; 1975: 1507.
- 67. Ebong, P.E., Atangwho, I.J., Eyong, E.U., Egbungand, G.E., Ikpeme, E.V. Effect of co-administration of extracts of *Vernonia amygdalina* and *Azadirachta indica* on lipid profile and oxidative stress in hepatocytes of normal and diabetic rats. Agriculture and Biology Journal of North America 2011; 7: 1087-95.
- 68. Tanko Y, Abdelaziz MM, Adelaiye AB, Fatihu MY, Musa KY. Effects of Hydromethanolic leave extract of *Indigofera pulchra* on blood glucose levels of normoglycemic and alloxan-induced diabetic Wistar rats. Int Journ Appl Res Nat Prod. 2009; 1: 13-8.
- Yeh G Y, Eisenberg D M, Kaptcuk T J and Philips R S. System review of herbs and dietary supplements for glycemic control in diabetes. Diabet Care. 2003; 26: 1277 – 94.

## ACKNOWLEDGEMENT: None

**CONFLICT OF INTEREST:** The authors declare no conflict of interest, financial or otherwise. **SOURCE OF FUNDING:** None.

SOURCE OF FUNDING: None.

**AVAILABILITY OF DATA AND MATERIALS:** Not applicable.

CONSENT FOR PUBLICATION: Not applicable.

ETHICS APPROVAL AND CONSENT TO **PARTICIPATE:** Not applicable.